

New insights into the synthesis and characterization of 2-methoxy-3-alkylpyrazines and their deuterated isotopologues[†]

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A previously described synthetic route for preparation of 2-methoxy-3-alkylpyrazines (MPs) based on condensation of glyoxal with an α -amino acid amide, followed by methylation with iodomethane yields 3-alkyl-1-methyl-1H-pyrazin-2-ones (*N*-methyl derivatives), rather than the designated 2-methoxy-3-alkylpyrazines (*O*-methyl derivatives). Despite similar nuclear magnetic resonance and mass spectral properties, gas chromatographic (GC) retention indices differ significantly, indicating chemical difference. With the example of 3-*sec*-butyl-1-methyl-1H-pyrazin-2-one and its 3-*sec*-butyl-1-[²H₃]methyl-1H-pyrazin-2-one isotopologue, the position of the methyl group introduced could be assigned unambiguously, using heteronuclear multiple bond correlation (HMBC) NMR experiments. For future characterization, the spectroscopic (NMR, EI⁺MS) as well as GC retention index data on two stationary phases of the most aroma relevant MPs and their deuterated isotopologues are summarized.

Keywords: methoxypyrazines; synthesis; characterization; NMR; GC-MS; linear retention indices

Introduction

2-Methoxy-3-alkylpyrazines (MPs; **4**) are a class of important and powerful flavor compounds. Owing to the extremely low flavor threshold values of some of the MPs, in particular those with iso-propyl (2-methoxy-3-iso-propylpyrazine (IPMP); **4a**), *sec*-butyl (2-methoxy-3-*sec*-butylpyrazine (SBMP); **4b**) and iso-butyl (2-methoxy-3-iso-butylpyrazine (IBMP); **4c**) as alkyl chain, they contribute to the aroma of important foodstuffs¹ such as bell pepper or peas^{2,3} and certain *Vitis vinifera* varieties.^{4,5}

Published methods for the synthesis of 2-methoxy-3-alkylpyrazines vary. A common approach to generate the heterocyclic ring system follows the route originally described by Jones⁶ for the synthesis of hydroxypyrazines **1**, in which an α -amino acid amide is condensed with a 1,2-dicarbonyl compound (Figure 1). Later, Karmas and Spoerri⁷ introduced the more accessible hydrohalides of the amides, and furthermore, described in detail the conversion of the preliminary hydroxypyrazines **1** to the 2-chloro derivatives **2** using phosphorous oxychloride⁸ (Figure 1, route A). The reaction of these 2-chloro derivatives with ethanolic sodium ethoxide, forming the ethyl ethers (ethoxy-pyrazines) has also been described in this early work and was used in subsequent studies for the synthesis of various 2-alkoxy-3-alkylpyrazines.^{9–12} Buttery and co-workers used an alternative approach for converting the preliminary hydroxypyrazines to 2-methoxy-3-alkylpyrazines **4** by derivatization with diazomethane^{2,13} (Figure 1, route B). However, they observed a by-product in a ratio of 2:1 (in favor of the by-product), which they described as 1,2-dihydro-3-alkyl-1-methyl-2-pyrazinone (or 3-alkyl-1-methyl-1H-pyrazin-2-one) **3**. The reason for this

result was supposed to be that hydroxypyrazines **1** exist largely in the pyrazinone **1*** form in neutral solution, which could be confirmed by IR¹⁴ or NMR¹⁵ experiments.

In a more recent publication, Gerritsma *et al.*¹⁶ described a method using sodium hydride and iodomethane to convert the originally formed hydroxypyrazines **1** to the targeted 2-methoxy-3-alkylpyrazines **4** (Figure 1, route C; later ascertained as a misconception, since this route leads to compounds **3** as outlined below). These authors also described the synthesis of the deuterated isotopologues (using [²H₃]iodomethane instead of iodomethane). Such compounds are important reference substances for trace level analysis of 2-methoxy-3-alkylpyrazines **4**,

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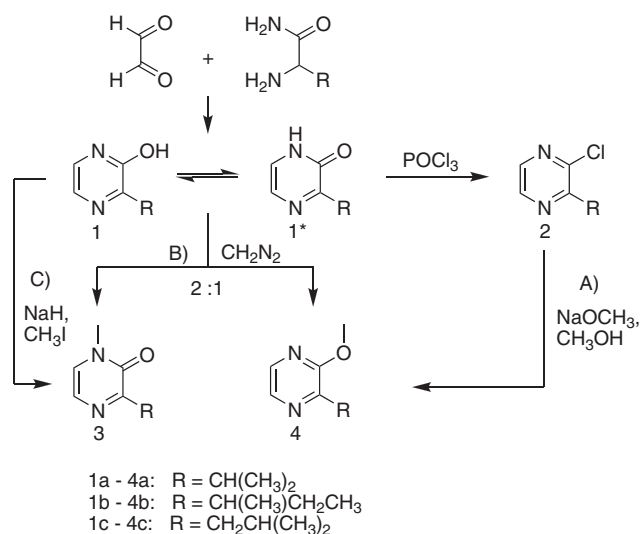


Figure 1. Routes to synthesize MPs as described in the literature.

following the approach of stable isotope dilution analysis (SIDA) for quantification.¹⁷

In the work described here, we synthesized the desired deuterated 2-[²H₃]methoxy-3-alkylpyrazines d₃-**4a-c** for wine aroma analysis. Considering the published methods described for MP synthesis, we first followed the method described by Gerritsma *et al.*¹⁶ (Figure 1, route C) as it seemed to be the most convenient approach. However, the products obtained showed different gas chromatographic (GC) properties compared with reference MPs **4a-c**. This paper clarifies some of the previously published results and summarizes the chromatographic and spectroscopic characterization of compounds obtained by the method described by Gerritsma *et al.*¹⁶ and MPs, respectively their deuterated isotopologues.

Results and discussion

In our hands, the deuterated derivatives obtained after the approach of Gerritsma *et al.*¹⁶ (Figure 1, route C) were not the designated MPs d₃-**4a-c**, as they showed considerably higher retention indices after GC analysis, than those for the commercial non-deuterated MP reference substances. We obtained substances, which could later be assigned to the structures of d₃-**3a-c**. Incorporation of deuterium into an organic molecule leads to a somewhat lowered retention index in gas liquid chromatography, despite its higher absolute molecular mass. This effect is thus called *inverse isotope effect*¹⁸ and has been attributed to the different binding-length of C-H and C-D as summarized by Matucha *et al.*¹⁹ The unexpected and drastically higher retention indices of more than 200 (on a 5% phenylmethylpolysiloxane stationary phase) or 600 (on a polyethylene glycol stationary phase) units for d₃-**3a-c** and **4a-c** were an indication for a differing chemical nature of the substances. Unfortunately, Gerritsma and co-workers provided only MS and NMR data, but MS data acquisition had obviously been achieved without prior GC separation. A full listing of retention and MS data can be found in the experimental supplementary section (Table S-1).

To verify synthesis procedures, deuterated sec-butyl pyrazine derivative was synthesized according to the originally described pathway (Figure 1, route A; d₃-**4b**) as well as the non-deuterated

compound (later assigned as **3b**) according to Gerritsma *et al.*¹⁶ (Figure 1, route C). GC and GC-MS data of the compound obtained from route A are consistent with those of the reference compound SBMP **4b** from Sigma-Aldrich (Table S-1). Considering earlier results,^{2,13} 1,2-dihydro-3-sec-butyl-1-methyl-2-pyrazinone (or 3-sec-butyl-1-methyl-1H-pyrazin-2-one) **3b** was considered a possible product obtained from route C. Direct comparison of the ¹H-NMR spectra of commercial SBMP **4b** and its deuterated isotopologue d₃-**4b** from route A and of **3b** and its deuterated isotopologue d₃-**3b** showed disappearance of the methyl-singlets at 3.96 and 3.49 ppm, respectively (full spectral listings are in the experimental supplementary information section). This chemical shift difference of almost 0.5 ppm and a remarkable upfield shift of the aromatic protons by about 1 ppm for **4b** clearly demonstrate the success of incorporating the CH₃-groups in different positions in compounds **4b** and **3b**. The pattern of the other signals of d₃-**4b** is comparable and in accordance with published data.^{11,12,20} In the ¹³C-NMR spectra of compound **3b** and of commercial MP **4b** (Figure S-2), the signal of the heteroatom bound methyl group is shifted about 16 ppm from 37.3 ppm in **3b** to 53.3 ppm in **4b**. The positions of this methyl groups were unambiguously assigned by heteronuclear correlation NMR (¹H-¹³C- and ¹H-¹⁵N-HMBC) experiments (Figures S-3 and S-4). This way, the ¹⁵N chemical shifts were estimated for both nitrogen atoms in **3b** with -217 and -54 ppm. This is comparable with literature values for the 1H-pyrazin-2-one with -198 and -36 ppm.²¹ It could be shown (Figure S-3) that the introduction of the methyl group (3.5 ppm) is at the nitrogen in the amide position (-217 ppm), whereas the CH hydrogen (3.3 ppm) of the sec-butyl group shows a coupling to the nitrogen in 4 position (-54 ppm) as expected. This demonstrates that the synthesis pathway suggested by Gerritsma *et al.* (route C) lead to the 3-alkyl-1-methyl-1H-pyrazin-2-ones **3**, and not to the desired MPs **4**. Such finding underlines the significance of simple retention index information for characterization of compounds amenable to GC, as spectral data can be similar.

Furthermore, a closer investigation of the mass spectra of deuterated **3b** (**3c**) and **4b** (**4c**) revealed a loss of 15 and 18 mass units (representing a methyl or a [²H₃]methyl group, respectively) which can also be used for differentiation of the two structures. Gerritsma *et al.*¹⁶ described the loss of either fragments from the molecular ion (*M* = 169) on the example of what they believed to be d₃-**4c**. In their mass spectral interpretation, this methyl group (or [²H₃]methyl group) loss could have come either from the methoxy position or the alkyl side chain, as they had found fragment ions at both *m/z* = 151 and 154 in the case of the deuterated compound. In our hands, such fragmentation was only observed for compounds d₃-**3b** and **c** (Figure S-5) and not for d₃-**4b** and **c** (Table S-1). Interestingly, in a thesis conducted at the same institution as Gerritsma's earlier work,²² Chen¹¹ observed this discrepancy in mass spectra from compounds synthesized according to route A (modified according to Masuda *et al.*¹⁰) as well. Unfortunately, no clear statement had been given that the earlier described synthesis obviously leads to wrong compounds **3** rather than the desired MPs **4**. This is the more surprising, as the same group recently published (as co-authors) a work on quantitative analysis of MPs in juice and wine using SIDA.¹² The original synthetic route of Karmas and Spoerri⁷ (A) was followed to obtain deuterated MPs d₃-**4**. However, commenting their earlier work¹⁶ had not been an option. With regard to the

shortcomings found in literature, full MS spectra are supplied in Figure S-5 for compounds d_3 -**3a-c** resulting from synthetic route C (shown in Figure 1) in addition to the text listings in Table S-1.

References

- [1] K. E. Murray, F. B. Whitfield, *J. Sci. Food Agric.* **1975**, *26*, 973–986.
- [2] R. G. Buttery, R. M. Seifert, D. G. Guadagni, L. C. Ling, *J. Agric. Food Chem.* **1969**, *17*, 1322–1327.
- [3] K. E. Murray, J. Shipton, F. B. Whitfield, *Chem. Industry* **1970**, *1970*, 897–898.
- [4] C. Bayonove, R. Cordonnier, Dubois, *C. R. Hebd. Seances Acad. Sci.* **1975**, *281*, 75–78.
- [5] O. P. H. Augustyn, A. Rapp, C. J. Van Wyk, *S. Afr. J. Enol. Vitic.* **1982**, *3*, 53–60.
- [6] R. G. Jones, *J. Am. Chem. Soc.* **1949**, *71*, 78–81.
- [7] G. Karmas, P. E. Spoerri, *J. Am. Chem. Soc.* **1952**, *74*, 1580–1584.
- [8] A. E. Erickson, P. E. Spoerri, *J. Am. Chem. Soc.* **1946**, *68*, 400–402.
- [9] G. Karmas, P. E. Spoerri, *J. Am. Chem. Soc.* **1956**, *78*, 4071–4077.
- [10] H. Masuda, M. Yoshida, T. Shibamoto, *J. Agric. Food Chem.* **1981**, *29*, 944–947.
- [11] X. Chen, Thesis (M.Sc.), Brock University, St. Catharines, Ont., Canada, **2005**.
- [12] Y. S. Kotseridis, M. Spink, I. D. Brindle, A. J. Blake, M. Sears, X. Chen, G. Soleas, D. Inglis, G. J. Pickering, *J. Chromatogr. A* **2008**, *1190*, 294–301.
- [13] R. M. Seifert, R. G. Buttery, D. G. Guadagni, D. R. Black, J. G. Harris, *J. Agric. Food Chem.* **1970**, *18*, 246–249.
- [14] H. Gainer, M. Kokorudz, W. K. Langdon, *J. Org. Chem.* **1961**, *26*, 2360–2363.
- [15] R. H. Cox, A. A. Bothner-By, *J. Phys. Chem.* **1968**, *72*, 1642–1645.
- [16] D. A. Gerritsma, I. D. Brindle, T. R. B. Jones, A. Capretta, *J. Label. Compd. Radiopharm.* **2003**, *46*, 243–253.
- [17] D. Rittenberg, G. L. Foster, *J. Biol. Chem.* **1940**, *133*, 733–744.
- [18] F. Bruner, G. P. Cartoni, A. Liberti, *Anal. Chem.* **1966**, *38*, 298–303.
- [19] M. Matucha, W. Jockisch, P. Verner, G. Anders, *J. Chromatogr. A* **1991**, *588*, 251–258.
- [20] R. L. N. Harris, M. J. Lacey, W. V. Brown, M. S. Allen, *Vitis* **1987**, *26*, 201–207.
- [21] S. Tobias, H. Günther, *Tetrahedron Lett.* **1982**, *23*, 4785–4788.
- [22] D. A. Gerritsma, Thesis (B.Sc.), Brock University, St. Catharines, Ont., Canada, **2001**.